

## iXCells Protocol

# IPSC-DERIVED CARDIOMYOCYTES MAINTENACE PROTOCOL

### Procedures:

1. Upon receipt of the frozen cells, it is recommended to thaw the cells and initiate the culture immediately in order to retain the highest cell viability.
2. To thaw the cells, put the vial in 37°C water bath with gentle agitation for ~2 minute. Keep the cap out of water to minimize the risk of contamination.
3. Transfer the cells into a 15 mL conical tube with 5 mL Cardiomyocytes Replating Medium (Cat# MD-0046). Centrifuge at 250 g for 5 minutes at room temperature.
4. Aspirate the supernatant, resuspend the cells in 1 mL Cardiomyocytes Replating Medium. Check cell number and viability. Dilute cell to a concentration of  $0.5 \times 10^6$ /mL and apply 1 mL/well into matrigel-coated 24 well plate (Cat# MD-0023). Put the plate into 37°C incubator overnight.
5. Next day, aspirate Cardiomyocytes Replating Medium, apply 1 mL Cardiomyocytes Maintenance Medium (Cat# MD-0045) to each well.
6. Change medium with Cardiomyocytes Maintenance Medium (Cat# MD-0045) every other day.

**Note:** Normally the cells will aggregate and start beating in 7~10 days after replating.

### Reagents/Media needed:

Reagent	Size	Catalog #
Cardiomyocytes Replating Medium	50 mL	MD-0046
Cardiomyocytes Maintenance Medium	100 mL	MD-0045
Matrigel-Coated Plate	each	MD-0023

### Disclaimers

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